

bKIT *Empetrum nigrum*

Real-Time PCR assay

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Empetrum nigrum

Empetrum nigrum, also called black crowberry, is a flowering plant of the northern hemisphere belonging to the Ericaceae family. Recently, it gained popularity due to its high content of flavonoids. These compounds are well known for their antioxidant, anti-inflammatory, and anticarcinogenic activities. Moreover, antibacterial and antifungal activities, as well a positive effect on lipid metabolism have been reported for fruits of *Empetrum nigrum*.¹

1: Jurikova T., Mlcek J., Skrovankova S., Balla S., Sochor J., Baron M., Sumczynski D.. "Black Crowberry (*Empetrum nigrum* L.) Flavonoids and Their Health Promoting Activity." *Molecules*. 2016 Dec 7;21(12). pii: E1685. Review. PubMed PMID:27941619; PubMed Central PMCID: PMC6272966.

Principle

Hydrolysis probe Real-Time PCR assay for the detection of *Empetrum nigrum*. The product is intended for research purpose only.

NHPRA validation

In the validation trials performed by NHPRA (Natural Health Product Research Alliance) the following species were tested: *Calluna vulgaris*, *Camellia sinensis*, *Ceratiola ericoides*, *Cirsium* spp., *Corema conradii*, *Curcuma longa*, *Empetrum atropurpureum*, *Ginkgo biloba*, *Kalmia angustifolia*, *Panax ginseng*, *Panax quinquefolius*, *Serenoa repens*, *Silybum marianum*, *Valeriana officinalis*, *Zingiber officinale*.

The validation trials highlighted the needing for further analyses to discriminate *Empetrum atropurpureum* and *Calluna vulgaris*.

bKIT *Empetrum nigrum* packaging

Part Number: bKTB-EN.01-50

qPCR Master Mix (1 tube)	50 tests
Positive Control (1 tube)	10 tests
Negative Control (1 tube)	10 tests

Part Number: bKTB-EN.01-100

qPCR Master Mix (2 tubes)	2 x 50 tests
Positive Control (1 tube)	20 tests
Negative Control (1 tube)	20 tests

Storage

-20°C. Avoid prolonged exposure to light and repeated freeze and thaw cycles.

Shelf life

If the bKIT is correctly stored, at constant-temperature freezer, its performance is guaranteed until the shelf life indicated on the tubes.

Additional material/reagents required

- DNA extraction tools and reagents
- Nuclease-free water
- Gloves
- Pipettes
- bCUBE® instrument or other Real-Time PCR instrument (*) with filters calibrated for FAM
- bCUBE® sample loading cartridge or, if using other Real-Time PCR instrument, samples loading support according to the instrument specifications.

(*) *This assay was especially developed to be used in association with the bCUBE® instrument, available from Hyris Ltd, but can be used also with any other compatible thermal cycler.*

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DNA extraction

Extract DNA from samples according to your usual protocol. If necessary, Hyris can recommend an extraction method. At this purpose, contact us at support@hyris.net.

Reaction Set-Up

- Thaw all the bKIT components by placing the tubes on ice.
- Gently mix the tubes content by swirling the tubes.
- Spin the tubes to let the content down.
- In new tubes, one for each sample, including the **Negative Control** and the **Positive Control** of the bKIT, prepare the Reaction Mix as shown in the table below:

Components	Volume
DNA sample or Positive Control or Negative Control	3 µl
qPCR Mastermix	17 µl
Total Volume	20 µl

Cartridge set-up

The procedure described is for the bCUBE® cartridge, but, if using a different Real-Time PCR instrument, the same procedure can be adopted for other loading sample supports with minor modifications.

1. Samples set-up

- Samples of the following types must be prepared to be loaded on the cartridge:
Positive Control for *Empetrum nigrum*
Negative Control for *Empetrum nigrum*
 Sample(s) to be tested.

2. Cartridge Loading

- Load the sample prepared as described in the previous section
- Carefully seal the cartridge with adhesive film in order to avoid any contamination.
- Load the cartridge onto the bCUBE®, then start the run.

Method set-up

Set up the run method using the following conditions, depending on the instrument you use.

1. On the bCUBE®

- Login on the bAPP.
- Set-up “New Analysis” and Select the “*Empetrum nigrum* 1.x” from the Global Recipes either on the bAPP.
- Define your wells types as follows (**Fig. 1**):
 “PosCtrl” for the well loaded with *Empetrum nigrum* **Positive Control**.
 “NegCtrl” for the well loaded with *Empetrum nigrum* **Negative Control**.
 “Sample” for the wells loaded with samples under analysis.



Fig 1. Cartridge set-up

An example of cartridge set-up on the bAPP for one replicate of a sample to be analyzed is shown.

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2. On a compatible Real-Time PCR instrument

Please, contact us for the protocol set-up on the instrument.

Results analysis

1. On the bCUBE®

a. The presence of the target *Empetrum nigrum* in the **Positive Control** or in the sample under analysis will generate an amplification curve (**Fig. 2**)

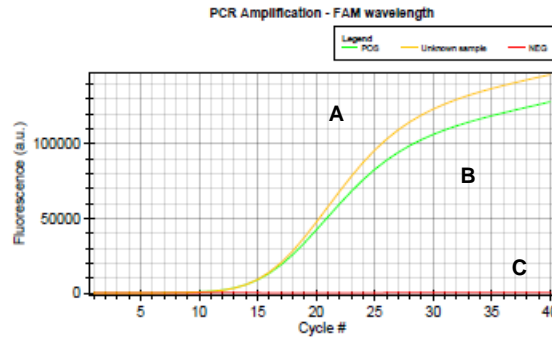


Fig.2. Amplification plot

In the plot, the amplification curve (a) of a *Empetrum nigrum* target containing sample (A), a **Positive Control** (B), and a **Negative Control** (C) are shown.

b. At the end of analysis each well will be labelled depending on the well type as described in the tables below:

Well	Possible labels	
Positive Control (PosCtrl)	OK	KO
Label meaning	Amplification curve present	Amplification curve absent

Well	Possible labels	
Negative Control (NegCtrl)	OK	KO
Label meaning	Amplification curve absent	Amplification curve present

Well	Possible labels		
Sample	Present	Absent	Indeterminate
Label meaning	<i>Empetrum nigrum</i> is present in the sample	<i>Empetrum nigrum</i> is absent from the sample	The test is not conclusive and should be repeated (**)

(**) If the Indeterminate classification persists, contact us.

c. On the bAPP, and on the pdf report of the analysis, the samples classification will be also shown (**Fig. 3**).

Results for target <i>Empetrum nigrum</i>		
Positive control (PosCtrl)		OK
Unknown sample (Sample)		Present
Negative control (NegCtrl)		OK

Fig.3. Analysis results table

The results table, as reported in the pdf report of the analysis, is shown.

2. On a compatible Real-Time PCR instrument

Please, contact us for results interpretation.

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Troubleshooting

1. Results show no amplification, or anomalous amplification curves

Possible causes	Corrective actions
Evaporation of the sample due to inadequate sealing of the plate/strips	Repeat the test using the appropriate tools to seal correctly the plate/strips
Consumables are not appropriate for the method	Repeat the test using consumables recommended by the supplier of the Real-Time PCR instrument
The quality of nucleic acid extracted is low	Repeat the extraction step. Ensure that the method of extraction has been performed correctly. In any doubt, contact us

2. No amplification curve is observed for the Positive Control

Possible causes	Corrective actions
The Positive Control provided with the assay was not added into the reaction well	Repeat the test. If the problem persists, contact us

3. An amplification curve with a specific melting peak is observed for the Negative Control

Possible causes	Corrective actions
Contamination of the Negative Control or the qPCR Master Mix with target-positive DNA	Repeat the test by applying appropriate quality procedures to prevent contamination. If the problem persists, contact us. Correctly seal the cartridge or plate/strips

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